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(54) Purified thermostable dna polymerase obtainable from pyrococcus species

Gereinigte thermostabile DNS-Polymerase aus Pyrococcus-Arten

DNA polymérase thermostable purifiée à obtenir des espèces de pyrococcus

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(73) Proprietor: NEW ENGLAND BIOLABS, INC.
 Beverly Massachusetts 01915 (US)

(72) Inventors:

- Comb, Donald G.
 Beverly, MA 01915 (US)
- Kucera, Rebecca
 Beverly, MA 01915 (US)
- Perler, Francine
 Brookline, MA 02146 (US)
- Kong, Hui-min
 Beverly, MA 01915 (US)
- Jack, William E.
 Wenham, MA 01984 (US)

(74) Representative: Davies, Jonathan Mark et al
 Reddle & Grose
 16 Theobalds Road
 London WC1X 8PL (GB)

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Description**FIELD OF THE INVENTION**

5 [0001] The present invention relates to an extremely thermostable enzyme. More specifically, it relates to a thermostable DNA polymerase obtainable from *Pyrococcus sp.*, an isolated DNA sequence coding for the polymerase, vectors and microbial hosts transformed with the DNA, a process for preparation of the polymerase and a process for increasing expression of the polymerase from a host.

BACKGROUND OF THE INVENTION

10 [0002] DNA polymerases are a family of enzymes involved in DNA repair and replication. Extensive research has been conducted on the isolation of DNA polymerases from mesophilic microorganisms such as *E. coli*. See, for example, Bessman, et al., *J. Biol. Chem.* (1957) 233:171-177 and Buttin and Kornberg *J. Biol. Chem.* (1966) 241:5419-5427.

15 [0003] Examples of DNA polymerases isolated from *E. coli* include *E. coli* DNA polymerase I, Klenow fragment of *E. coli* DNA polymerase I and T4 DNA polymerase. These enzymes have a variety of uses in recombinant DNA technology including, for example, labelling of DNA by nick translation, second-strand cDNA synthesis in cDNA cloning, and DNA sequencing. See Maniatis, et al., *Molecular Cloning: A Laboratory Manual* (1982).

20 [0004] Recently, U.S. Patent Nos. 4,683,195, 4,683,202 and 4,800,159 disclosed the use of the above enzymes in a process for amplifying, detecting, and/or cloning nucleic acid sequences. This process, commonly referred to as polymerase chain reaction (PCR), involves the use of a polymerase, two primers and nucleotide triphosphates to amplify existing nucleic acid sequences.

25 [0005] Some of the DNA polymerases discussed above possess a 3'-5' exonuclease activity which provides a proof-reading function that gives DNA replication much higher fidelity than it would have if synthesis were the result of only a one base-pairing selection step. Brutlag, D. and Kornberg, A., *J. Biol. Chem.* (1972) 247:241-248. DNA polymerases with 3'-5' proofreading exonuclease activity have a substantially lower base incorporation error rate when compared with a DNA polymerase without proofreading activity. Chang, L.M.S., *J. Biol. Chem.* (1977) 252:1873-1880.

30 [0006] Research has also been conducted on the isolation and purification of DNA polymerases from thermophiles, such as *Thermus aquaticus*. Chien, A., et al. *J. Bacteriol.* (1976) 127:1550-1557, discloses the isolation and purification of a DNA polymerase with a temperature optimum of 80°C from *T. aquaticus* YT1 strain. The Chien, et al., purification procedure involves a four-step process. These steps involve preparation of crude extract, DEAE-Sephadex chromatography, phosphocellulose chromatography, and chromatography on DNA cellulose. Kaledin, et al., *Biokhimiya* (1980) 45:644-651 also discloses the isolation and purification of a DNA polymerase from cells of *T. aquaticus* YT1 strain. The Kaledin, et al. purification procedure involves a six-step process. These steps involve isolation of crude extract, ammonium sulfate precipitation, DEAE-cellulose chromatography, fractionation on hydroxyapatite, fractionation on DEAE-cellulose, and chromatography on single-strand DNA-cellulose.

35 [0007] United States Patent No. 4,889,818 discloses a purified thermostable DNA polymerase from *T. aquaticus*, Taq polymerase, having a molecular weight of about 86,000 to 90,000 daltons prepared by a process substantially identical to the process of Kaledin with the addition of the substitution of a phosphocellulose chromatography step in lieu of chromatography on single-strand DNA-cellulose. In addition, European Patent Application 0258017 discloses Taq polymerase as the preferred enzyme for use in the PCR process discussed above.

40 [0008] Research has indicated that while the Taq DNA polymerase has a 5'-3' polymerase-dependent exonuclease function, the Taq DNA polymerase does not possess a 3'-5' proofreading exonuclease function. Lawyer, F.C., et al. *J. Biol. Chem.* (1989) 264:11, p. 6427-6437. Bernad, A., et al. *Cell* (1989) 59:219. As a result, Taq DNA polymerase is prone to base incorporation errors, making its use in certain applications undesirable. For example, attempting to clone an amplified gene is problematic since any one copy of the gene may contain an error due to a random misincorporation event. Depending on where in the replication cycle that error occurs (e.g., in an early replication cycle), the entire DNA amplified could contain the erroneously incorporated base, thus, giving rise to a mutated gene product. Furthermore, research has indicated that Taq DNA polymerase has a thermal stability of not more than several minutes at 100°C. Therefore, a DNA polymerase with much higher thermal stability and an associated 3' to 5' exonuclease proofreading activity is urgently needed by the scientific community. One such enzyme, the DNA polymerase from *Thermococcus litoralis*, an archaebacterium that grows at temperatures close to 100°C near submarine thermal vents, has recently been isolated and cloned in *E. coli*.

45 [0009] However, there is still a desire in the art to obtain and produce a purified DNA polymerase having 3'-5' proof-reading exonuclease activity with even greater thermostability than DNA polymerases currently available. The availability of such an enzyme would improve the DNA polymerase processes described above. In addition, it would be useful if such a DNA polymerase were produced by recombinant DNA techniques. This would allow for the production of highly pure commercial quantities of the polymerase.

SUMMARY OF THE INVENTION

[0010] In accordance with the present invention, there is provided a purified thermostable enzyme obtainable from *Pyrococcus sp.* which catalyzes the polymerization of DNA wherein said enzyme is obtainable from *E.coli* NEB #720 (ATCC # 68723). The thermostable enzyme obtainable from *Pyrococcus sp.*, an archaebacterium isolated from a submarine thermal vent at 2010 meters, is a DNA polymerase which has an apparent molecular weight of about 92,000-97,000 daltons, a half-life of about 8 hours at 100°C and a half-life of 23 hours at 95°C.

[0011] The DNA encoding the 92,000-97,000 daltons thermostable DNA polymerase obtainable from *Pyrococcus sp.* has been isolated and provides a means to obtain the thermostable enzyme of the present invention.

[0012] The *Pyrococcus sp.* DNA polymerase possesses 3'-5' proofreading exonuclease activity. In accordance with the present invention, it has been found that the 3'-5' proofreading activity of the *Pyrococcus sp.* DNA polymerase is 2.5 times that of the DNA polymerase obtainable from *T. litoralis*. As a result, *Pyrococcus sp.* DNA polymerase should have a much higher fidelity than a thermostable polymerase with no 3'-5' proofreading exonuclease function, such as Taq polymerase, and may have a greater proofreading activity than *T. litoralis* DNA polymerase. In addition, the *Pyrococcus sp.* DNA polymerase has a substantially greater thermal stability or half life at temperatures from 96°C to 103°C than the Taq polymerase.

BRIEF DESCRIPTION OF DRAWINGS

[0013] FIG. 1A - is the ethidium bromide stained agarose gel of *Pyrococcus sp.* DNA cut with EcoR I (lane 3), BarnH I (lane 4) and Hind III (lane 5). Lane 1 is λDNA cut with Hind III as markers and lane 2 is pBR322 as a marker.

[0014] FIG. 1B - is an autoradiography of a Southern Hybridization of the same gel in 1A. The ³²P-DNA probe was prepared from a 1.3 Kb Eco RI fragment that encodes the amino terminal portion of the *Thermococcus litoralis* DNA polymerase. Note that the BarnH I cut *Pyrococcus sp.* DNA gives a single band of about 4-5 Kb with the probe. The fact that the 23 Kd band of Hind III cut λDNA shows up on the film is due to nonspecific hybridization to the large amount of DNA present in that band. The fact that the plasmid pBR322 lights up is due to homologous sequences in the probe.

[0015] FIG. 2 - is a restriction site map of the 4.8 Kb BarnH I fragment containing the gene encoding the *Pyrococcus sp.* DNA polymerase in the pUC19 plasmid of *E. coli* 2207 (NEB#720).

[0016] FIG. 3 - is a photograph of the Coomassie Blue stained gel from which the approximate molecular weight of the *Pyrococcus sp.* DNA polymerase was determined. The arrow beside lane 1 indicates the polymerase band. Lane 2 contains the indicated molecular weight standards.

[0017] FIG. 4 - is a photograph of the western blot comparing *Thermococcus litoralis* DNA polymerase with *Pyrococcus sp.* DNA polymerase. Lane 1, Protein standards; Lane 2, 50 μg of a crude extract of *Thermococcus litoralis*; Lane 3, 2.0 μg of purified recombinant *Pyrococcus sp.* DNA polymerase. The arrow indicates the position of the polymerase I.

[0018] FIG. 5 - is a comparison of the polymerizing and exonuclease function of *Thermococcus litoralis* DNA polymerase and the *Pyrococcus sp.* DNA polymerase.

[0019] FIG. 6 - shows the heat stability at 95°C (O) and 100°C (O) of the recombinant *Pyrococcus sp.* DNA polymerase.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0020] The present invention relates to a thermostable enzyme which is a DNA polymerase obtainable from *Pyrococcus sp.* strain GB-D. *Pyrococcus sp.* was isolated from a submarine thermal vent in the Sea of Cortez at a depth of 2,010 meters by Holger Jannasch using the deep sea submersible submarine Alvin of the Woods Hole Oceanographic Institute. A sample of *Pyrococcus sp.* strain GB-D (Archaebacterium, NEB#732) was deposited with the American Type Culture Collection on October 1, 1991, under the terms of the Budapest Treaty, and received ATCC Accession No. 55239.

[0021] This organism, *Pyrococcus sp.*, is an extremely thermophilic, sulfur metabolizing, archaebacterium, with a growth range between 65°C and 103°C.

[0022] For recovering the native protein, *Pyrococcus sp.* may be grown using any suitable technique, such as the technique described by Belkin, et al., *Arch. Microbiol.* (1985) 142:181-186. Briefly, the cells are grown in the media described in Belkin, et al., *supra*, containing 10 mg/ml of sulfur and 0.01 M Cysteine in 15 ml screw cap tubes at 95°C for 2 days. When larger amounts of cells are required, 1 liter screw cap bottles are used, and after sterilization, are inoculated with a fresh 10 ml culture and grown at 90°C-95°C for 2 days.

[0023] After cell growth, one preferred method for isolation and purification of the enzyme is accomplished using the multi-step process as follows: First, the cells, if frozen, are thawed, suspended in a suitable buffer such as buffer A (10

mM KP04 buffer, pH 7.4; 1.0 mM EDTA, 1.0 mM beta-mercaptoethanol), sonicated and centrifuged. The supernatant is then passed through a column which has a high affinity for proteins that bind to nucleic acids such as Affigel blue column (Biorad). The nucleic acids present in supernatant solution of *Pyrococcus sp.* and many of the proteins pass through the column and are thereby removed by washing the column with several column volumes of low salt buffer at pH of about 7.0. After washing, the enzyme is eluted with a linear gradient such as 0.1 to 2.0 M NaCl buffer A. The peak DNA polymerase activity is dialyzed and applied to phosphocellulose column. The column is washed and the enzyme activity eluted with a linear gradient such as 0.1 to 1.0 M NaCl in buffer A. The peak DNA polymerase activity is dialyzed and applied to an HPLC mono-S column (cation exchanger). The enzyme is eluted with a linear gradient such as 0.05 to 1.0 M NaCl in buffer A. The enzyme is about 50% pure at this stage.

[0024] The apparent molecular weight of the DNA polymerase obtainable from *Pyrococcus sp.* is between about 92,000 to 97,000 daltons when compared with protein standards of known molecular weight, such as phosphorylase B assigned a molecular weight of 97,400 daltons. It should be understood, however, that as a protein from an extreme thermophile, *Pyrococcus sp.* DNA polymerase may electrophorese at an aberrant relative molecular weight due to failure to completely denature or other intrinsic properties. The exact molecular weight of the thermostable enzyme of the present invention may be determined from the coding sequence of the *Pyrococcus sp.* DNA polymerase gene. The molecular weight of the eluted product may be determined by any technique, for example, by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) using protein molecular weight markers.

[0025] Polymerase activity is preferably measured by the incorporation of radioactively labeled deoxynucleotides into DNase-treated, or activated, DNA; following subsequent separation of the unincorporated deoxynucleotides from the DNA substrate, polymerase activity is proportional to the amount of radioactivity in the acid-insoluble fraction comprising the DNA. Lehman, I.R., et al., *J. Biol. Chem.* (1958) 233:163.

[0026] The half-life of the DNA polymerase of the present invention at 100°C is about 8 hours and about 23 hours at 95°C. The thermal stability or half-life of the DNA polymerase can be determined by preincubating the enzyme at the temperature of interest in the presence of the reaction buffer containing BSA. At predetermined time intervals, ranging up to 12 hours, small aliquots are removed, and added to reaction buffer containing DNA substrate, dNTPs, and BSA and assayed for polymerase activity using the method described above.

[0027] The thermostable enzyme of this invention may be produced by recombinant DNA techniques, as the gene encoding this enzyme has been cloned from *Pyrococcus sp.* genomic DNA. A partial DNA sequence of this gene is set forth in the Sequence Listing as SEQ ID NO:1. The complete coding sequence for the *Pyrococcus sp.* DNA polymerase can be derived from an approximately 5 Kb BamHI restriction fragment in pUC19 in *E. coli*, NEB#720. This *E. coli* strain was deposited with the American Type Culture Collection (ATCC) on October 1, 1991, under the terms of the Budapest Treaty and received Accession No. ATCC 68723.

Cloning of the *Pyrococcus sp.* DNA Polymerase

[0028] The production of a recombinant form of the *Pyrococcus sp.* DNA polymerase generally includes the following steps: DNA is isolated which encodes the active form of the polymerase, either in its native form or as a fusion with other sequences which may or may not be cleaved away from the native form of the polymerase and which may or may not effect polymerase activity. Next, the gene may or may not be operably linked to appropriate control sequences for expression in either prokaryotic or eukaryotic host/vector systems. The vector preferably encodes all functions required for transformation and maintenance in a suitable host, and may encode selectable markers and/or control sequences for *Pyrococcus sp.* polymerase expression. The vector is used to transform a suitable host. Active recombinant thermostable polymerase can be produced by transformed host cultures either continuously or after induction of expression. Active thermostable polymerase can be recovered either from within host cells or from the culture media if the protein is secreted through the cell membrane.

[0029] While each of the above steps can be accomplished in a number of ways, it has been found that for cloning the DNA encoding *Pyrococcus sp.* DNA polymerase, expression of the polymerase from its own control sequences in *E. coli* results in high levels of gene expression.

[0030] It has been discovered that at least one intervening sequence is present within the *Pyrococcus* DNA polymerase gene. By comparison of the *Pyrococcus species* polymerase DNA and protein sequences of the present invention with the *Thermococcus litoralis* DNA polymerase DNA and protein, it was determined that the *Pyrococcus* intervening sequence is within the conserved pol α motif region III, and begins at nucleotide 1839 of SEQ ID NO:1.

Cloning Vectors

[0031] Vectors useful in practicing the present invention should provide varying degrees of controlled expression of *Pyrococcus sp.* polymerase by providing some or all of the following control features: (1) promoters or sites of initiation of transcription, either directly adjacent to the start of the polymerase or as fusion proteins, (2) operators which could

be used to turn gene expression on or off, (3) ribosome binding sites for improved translation, and (4) transcription or translation termination sites for improved stability. Appropriate vectors used in cloning and expression of *Pyrococcus* sp. polymerase include, for example, phage and plasmids. Examples of phage include λ gt11 (Promega), λ DASH (Stratagene) λ ZapII (Stratagene). Examples of plasmids include pUC19, pBR322, pBluescript (Stratagene), pSP73 (Promega), pGW7 (ATCC No. 40166), pET3A (Rosenberg, et al., *Gene*, (1987) 56:125-135), and pET11C (Methods in Enzymology (1990) 185:60-89).

Transformation and Infection

[0032] Standard protocols exist for transformation, phage infection and cell culture. Maniatis, et al., *Molecular Cloning: A Laboratory Manual* (1982). Of the numerous *E. coli* strains which can be used for plasmid transformation, the preferred strains include JM101 (ATCC No. 33876), XL1 (Stratagene), RRI (ATCC No. 31343), and BL21(DE3) *plysS* (*Methods in Enzymology* (1990), *supra*). *E. coli* strain XL1, ER1578 and ER1458 (Raleigh, et al., *N.A. Research* (1988) 16:1563-1575) are among the strains that can be used for λ phage, and Y1089 can be used for λ gt11 lysogeny. When preparing transient lysogens in Y1089 (Arasu, et al., *Experimental Parasitology* (1987) 64:281-289), a culture is infected with λ gt11 recombinant phage either by a single large dose of phage or by co-culturing with a lytic host. The infected Y1089 cells are preferably grown at 37°C in the presence of the inducer IPTG resulting in buildup of recombinant protein within the lysis-defective host/phage system.

Construction of Genomic DNA Library and Screening for Thermostable Polymerase

[0033] Probing a *Pyrococcus* genomic DNA library by cross-hybridization with radioactive probes prepared from the DNA polymerase gene of *Thermococcus litoralis* allowed for the identification and isolation of the *Pyrococcus* sp. DNA polymerase gene.

[0034] *Pyrococcus* sp. DNA can be isolated using the method described by Maniatis, et al., *Molecular Cloning: A Laboratory Manual* (1982).

[0035] The *Pyrococcus* sp. DNA once isolated can be used to construct genomic libraries as either random fragments or restriction enzyme fragments. The latter approach is preferred. Preferably, BamH I partials are prepared from *Pyrococcus* sp. genomic DNA using standard DNA restriction techniques such as described in Maniatis, et al., *supra*. Other restriction enzymes such as Sal I and XbaI can also be used.

[0036] Although methods are available to screen both plasmids and phage using antibodies or DNA probes (Young and Davis, *PNAS* (1983) 80:1194-1198; Maniatis et al., *supra*) it has been found that phage systems tend to work better and are therefore preferred for the first libraries.

[0037] Genomic libraries can be screened using the colony or plaque hybridization procedure (Maniatis, et al., *supra*) or using the antibody plaque reactivity (Young and Davis, *supra*). In the colony or plaque hybridization procedure, probes are formed by standard methods of labeling, for example, random priming or nick translation of a polymerase gene from a related organism, for example, *T. litoralis*. Maniatis, et al., *supra*. The genomic library is hybridized with labeled probe under conditions which depend on the stringency desired.

[0038] When genomic expression libraries are screened using the antibody/plaque procedure, since it is uncertain whether *Pyrococcus* sp. control regions function in *E. coli*, phage vectors which supply all necessary expression control regions such as λ gt11 and λ Zap II are preferred for antibody screening. By cloning *Pyrococcus* sp. DNA into the BamH I site of λ DASH or the EcoR I site of λ gt11, *Pyrococcus* sp. polymerase may be expressed either as a fusion protein with beta-galactosidase in λ gt11 or from its own endogenous promoter.

[0039] Once formed, the expression libraries can be screened with anti-*Pyrococcus* sp. DNA polymerase antiserum or if not available, by antibody against the DNA polymerase of a closely related organism (i.e. *Thermococcus litoralis*, another extreme thermophile) using standard antibody plaque hybridization procedures such as those described by Young and David, *PNAS* (1983), *supra*.

[0040] Using either procedure, the *Pyrococcus* sp. DNA polymerase DNA, coding for part or the whole gene, once identified can then be subcloned in, for example, pBR322, pBluescript, M13 or pUC19. If desired, the DNA sequence can be determined by, for example, the Sanger dideoxy chain-terminating method (Sanger, F., Nicklen, S. & Coulson, A.R. *PNAS* (1977) 74:5463-5467).

Identification of DNA Encoding and Expression of the *Pyrococcus Species* DNA Polymerase.

[0041] Several methods exist for determining that the DNA sequence coding for the *Pyrococcus* sp. DNA polymerase has been obtained. These include, for example, comparing the actual or deduced amino-terminal sequence of the protein encoded by the recombinant DNA to the native protein, or determining whether the recombinant DNA produces a protein which binds antibody specific for native *Pyrococcus* sp. DNA polymerase. In addition, research by Wang, et

al., *FASEB Journal* (1989) 3:14-21 suggests that certain regions of DNA polymerase sequences from the polymerase α family are highly conserved among many species as well as a separate group of conserved regions in Pol-like polymerases. As a result, by comparing the predicted amino acid sequence of the cloned gene with the amino acid sequence of known DNA polymerases, such as human DNA polymerase α and *E. coli* DNA polymerase I, the identification of these islands of homology provides strong evidence that the recombinant DNA indeed encodes a DNA polymerase.

[0042] As noted above, it has been found that DNA coding for *Pyrococcus* sp. DNA polymerase contains an intron or intervening sequence within the pol α motif region III. Therefore, as one means to increase overexpression in host cells such as *E. coli*, the DNA sequence coding for the intron can be deleted. There are a number of approaches known in the art which can be used to delete DNA sequences and therefore splice out an intron *in-vitro*. One method involves identifying unique restriction enzyme sites in the coding region which are near the splice junction or area to be deleted. A duplex oligomer is synthesized to bridge the gap between the 2 restriction fragments. A 3-part ligation consisting of the amino end restriction fragment, the bridging oligo and the carboxy end restriction fragment yields an intact gene with the intron deleted.

[0043] Another method is a modification of the above-described method. The majority of the intron is deleted by cutting with restriction enzymes with unique sites within the intron, but close to the coding sequence border. The linear plasmid containing a deletion of the majority of the intron is ligated together. Single strand phage are generated from the pBluescript vector recombinant by superinfection with the f1 helper phage IR1. A single strand oligomer is synthesized with the desired final sequence and is annealed to the partially deleted intron phage DNA. The remainder of the intron is thus looped out. By producing the original phage in *E. coli* strain CJ236 the Kunkel method of mutagenesis (*Methods in Enzymology*, 154:367 (1987)) can be used to select for the full deleted intron constructs.

[0044] Yet another method which can be used to delete the intron uses DNA amplification. See, for example, Maniatis, et al., *Molecular Cloning: A Laboratory Manual*, (1989), Vol. 2, 2nd edition. Briefly, primers are generated to amplify and subsequently join the amino and carboxyl halves of the gene.

[0045] When an intron is deleted *in-vitro*, using the methods discussed above, the native splice junction may be unknown. Accordingly, one skilled in the art would predict that several possible artificial splice junctions exist that would result in the production of an active enzyme.

[0046] Once identified, the DNA sequence coding for the *Pyrococcus* sp. DNA polymerase, with or without the intron deleted, can be cloned into an appropriate expression vector such as a plasmid derived from *E. coli*, for example, pET3A, pBluescript or pUC19, the plasmids derived from the *Bacillus subtilis* such as pUB110, pTP5 and pC194, plasmids derived from yeast such as pSH19 and pSH15, bacteriophage such as λ phage, bacteria such as *Agrobacterium tumefaciens*, animal viruses such as retroviruses and insect viruses such as Baculovirus.

[0047] The recombinant vector is introduced into the appropriate host using standard techniques for transformation and phage infection. For example, the calcium chloride method, as described by Cohen, S.N., *PNAS* (1972) 69:2110 is used for *E. coli*. The transformation of *Bacillus* is carried out according to the method of Chang, S., et al., *Molecular and General Genetics* (1979) 168:111. Transformation of yeast is carried out according to the method of Parent, et al., *Yeast* (1985) 1:83-138. Certain plant cells can be transformed with *Agrobacterium tumefaciens*, according to the method described by Shaw, C.H., et al., *Gene* (1983) 23:315. Transformation of animal cells is carried out according to, for example, the method described in *Virology* (1973) 52:456. Transformation of insect cells with Baculovirus is carried out according to, for example, the method described in *Biotechnology* (1988) 6:47.

[0048] The transformants are cultivated, depending on the host cell used, using standard techniques appropriate to such cells. For example, for cultivating *E. coli*, cells are grown in LB media (Maniatis, *supra*) at 30°C to 42°C to mid log or stationary phase.

[0049] The *Pyrococcus* sp. DNA polymerase can be isolated and purified from a culture of transformed host cells, for example, by either extraction from cultured cells or the culture solution.

[0050] When the *Pyrococcus* sp. DNA polymerase is to be extracted from a cultured cell, the cells are collected after cultivation by methods known in the art, for example, centrifugation. Then, the collected cells are suspended in an appropriate buffer solution and disrupted by ultrasonic treatment, lysozyme and/or freeze-thawing. A crude extract containing the *Pyrococcus* sp. DNA polymerase is obtained by centrifugation and/or filtration.

[0051] When the *Pyrococcus* sp. DNA polymerase is secreted into the culture solution, i.e., alone or as a fusion protein with a secreted protein such as maltose binding protein, the supernatant is separated from the cells by methods known in the art, for example, centrifugation.

[0052] The separation and purification of the *Pyrococcus* sp. DNA polymerase contained in the culture supernatant or the cell extract can be performed by the method described above, or by appropriate combinations of known separating and purifying methods. These methods include, for example, methods utilizing solubility such as salt precipitation and solvent precipitation, methods utilizing the difference in molecular weight such as dialysis; ultra-filtration, gel filtration, and SDS-polyacrylamide gel electrophoresis, methods utilizing a difference in electric charge such as ion-exchange column chromatography, methods utilizing specific affinity such as affinity chromatography, methods utilizing

a difference in hydrophobicity such as reverse-phase high performance liquid chromatography and methods utilizing a difference in isoelectric point such as isoelectric focusing electrophoresis.

[0053] One preferred method for isolating and purification of the recombinant enzyme is accomplished using the multi-stage process as follows: First, the cells, if frozen are thawed, suspended in a suitable buffer such as Buffer A (100 mM NaCl, 25 mM Tris pH 7.5, 0.1 mM EDTA, 10% glycerol, 0.05% Triton X-100), lysed and centrifuged. The clarified crude extract is then heated to 75°C for approximately 30 minutes. The denatured proteins are removed by centrifugation. The supernatant is then passed through a column that has high affinity for proteins that bind to nucleic acids such as Affigel Blue column (Biorad). The nucleic acids present in the supernatant solution and many of proteins pass through the column and are thereby removed by washing the column with several column volumes with low-salt buffer at pH of about 7.0. After washing, the enzyme is eluted with a linear gradient such as 0.1 M to 1.5 M NaCl Buffer A. The active fractions are pooled, dialyzed and applied to a phosphocellulose column. The column is washed and DNA polymerase activity eluted with a linear gradient of 0.1 to 1.0 M NaCl in Buffer B (100 M NaCl, 15 mM KP04, 0.1 mM EDTA, 10% glycerol, 0.05% Triton X-100, pH 6.8). The fractions are collected and BSA is added to each fraction. The fractions with DNA polymerase activity are pooled. The *Pyrococcus* sp. DNA polymerase obtained may be further purified using the standard product purification techniques discussed above.

Stabilization and Use of the *Pyrococcus* species DNA Polymerase.

[0054] For long-term storage, the thermostable enzyme of the present invention is stored in the following buffer: 0.05 M NaCl, 0.01 M KP04 (pH 7.4), 0.1 mM EDTA and 50% glycerol at -20°C.

[0055] The *Pyrococcus* sp. DNA polymerase of the present invention may be used for any purpose in which such an enzyme is necessary or desirable. For example, in recombinant DNA technology including, second-strand cDNA synthesis in cDNA cloning, and DNA sequencing. See Maniatis, et al., *supra*.

[0056] The *Pyrococcus* sp. DNA polymerase of the present invention may be modified chemically or genetically to inactivate the 3'-5' exonuclease function and used for any purpose in which such a modified enzyme is desirable, e.g., DNA sequencing.

[0057] For example, genetically modified *Pyrococcus* sp. DNA polymerase may be isolated by randomly mutagenizing the *Pyrococcus* sp. DNA polymerase gene and then screening for those mutants that have lost exonuclease activity, without loss of polymerase activity. Alternatively, genetically modified *Pyrococcus* sp. DNA polymerase is preferably isolated using the site-directed mutagenesis technique described in Kunkel, T.A., *PNAS* (1985) 82:488-492.

[0058] In addition, the *Pyrococcus* sp. DNA polymerase of the present invention may also be used to amplify DNA, e.g., by the procedure disclosed in U.S. Patent Nos. 4,683,195, 4,683,202 and 4,800,159.

[0059] The following examples are given to illustrate embodiments of the present invention as it is presently preferred to practice. It will be understood that the examples are illustrative, and that the invention is not to be considered as restricted except as indicated in the appended claims.

EXAMPLE I

PURIFICATION OF A THERMOSTABLE DNA POLYMERASE FROM *PYROCOCCUS* SPECIES

[0060] *Pyrococcus* sp. strain GB-D (ATCC No. 55239) was grown in the media described by Belkin, et al., *supra*, containing 10 g/l of elemental sulfur in 8 one liter bottles at 94°C for two days. The cells were cooled to room temperature, separated from unused sulfur by decanting and collected by centrifugation and stored at -70°C. The yield of cells was 1.4 g per liter.

[0061] 11.5 g of cells obtained as described above, were suspended in 28 ml of buffer A (10 mM KP04 buffer, pH 7.4; 0.1 mM EDTA, 1.0 mM beta-mercaptoethanol) containing 0.1 M NaCl and sonicated for 5 minutes at 4°C. The lysate was centrifuged at 15,000 g for 30 minutes at 4°C. The supernatant solution was passed through a 18 ml Affigel blue column (Biorad). The column was then washed with 50 ml of buffer A containing 0.1 M NaCl. The column was eluted with a 300 ml linear gradient from 0.1 to 2.0 M NaCl in buffer A. The DNA polymerase eluted as a single peak at approximately 1.3 M NaCl and represented 90% of the activity applied. The peak activity of DNA polymerase (25 ml) was dialyzed against 1 liter of buffer A containing 100 mM NaCl, and then applied to 15 ml Phosphocellulose column, equilibrated with buffer A containing 100 mM NaCl. The column was washed with 50 ml of buffer A containing 100 mM NaCl, and the enzyme activity was eluted with 200 ml linear gradient of 0.1 to 1.0 M NaCl in buffer A. The activity eluted as a single peak at 0.6 M NaCl and represented 70% of the activity applied. The pooled activity (42 ml) was dialyzed against 500 ml of buffer A and applied to a 25 ml DEAE column. The column was washed with 50 ml of buffer A containing 0.1 M NaCl, and two-thirds of the enzyme activity passed through the column. The active fractions were pooled (30 ml) and applied to an 1.0 ml HPLC mono-S column (Pharmacia) and eluted with a 100 ml linear gradient in buffer A from 0.05 to 1.0 M NaCl. The activity eluted as a single peak at 0.22 M NaCl and represented 80%

of the activity applied.

[0062] Purified *Pyrococcus* sp. polymerase was electrophoresed in SDS 10-20% polyacrylamide gel and stained with either Coomassie Blue or the colloidal stain (ISS Problue) previously described to detect protein. A faintly staining protein band was seen at about 92,000 to 97,000 daltons; this molecular weight determination was obtained by comparison on the same gel to the migration of the following marker proteins (Bethesda Research Laboratories): myosin, 200,000 daltons; phosphorylase B, 97,400 daltons; BSA, 68,000 daltons; ovalbumin, 43,000 daltons, carbonic anhydrase 29,000 daltons; b-lactoglobulin, 18,400 daltons; lysozyme 14,300 daltons.

EXAMPLE II

CLONING OF *PYROCOCCUS SPECIES* DNA POLYMERASE GENE

[0063] Cross hybridization of a *Pyrococcus* genomic DNA library using radioactive probes prepared from the DNA polymerase gene of *Thermococcus litoralis* allowed for the identification and isolation of a DNA encoding the *Pyrococcus* DNA polymerase. This was accomplished as set forth below.

[0064] In order to determine which restriction enzymes would be most useful in preparation of the *Pyrococcus* genomic library, *Pyrococcus* sp. DNA was cut to completion with Eco RI, BamHI and HindIII. This DNA was subject to agarose gel electrophoresis (Figure 1A) and Southern hybridization (Figure 1B) using a DNA probe prepared as follows. A reaction mixture containing approximately 1 µg of the first EcoRI fragment of the *T. litoralis* DNA polymerase gene (bp 1-1274, obtainable from bacteriophage NEB#618, ATCC No. 40794) as a template in a commercial random priming kit (New England Biolabs, Inc.) was incubated for 1 hour at 37°C to produce a DNA probe of high specific activity. The probe was hybridized to *Pyrococcus* sp. DNA prepared above under moderately stringent conditions (Hybridization: overnight at 50°C, 4X SET, 0.1M sodium phosphate, pH 7, 0.1% Na pyrophosphate, 0.1% SDS, 1X Denhardt's solution; Wash Conditions: wash 3X 20-30 min. 45°C, 0.1X SET, 0.1M sodium phosphate, pH 7, 0.1% Na pyrophosphate, 0.1% SDS. Maniatis, et al., *supra*). A single major band at about 5 Kb was detected in BamH I cut *Pyrococcus* DNA. EcoR I and Hind III gave multiple bands with this probe, indicating that these enzymes cut within the *Pyrococcus* polymerase gene.

[0065] Based on these results, a BamHI genomic library was constructed using the phage vector λDASH (Stratagene). Partial and complete BamHI digests of *Pyrococcus* DNA were prepared. A mixture of the partial and completely BamHI digested DNA was ligated into the BamHI site of λDASH. The ligation mixture was packaged using Gigapack Gold (Stratagene) according to manufacturer's instructions and plated on *E. coli* ER1458. The packaged phage library contained 1 x 10⁶ phage per ml.

[0066] ³²P-labelled DNA probes of 3 fragments (SEQ ID NO:2, bp 1-1274, 1656-2660, and 3069-3737) of the *T. litoralis* DNA polymerase gene (obtainable from NEB#619, ATCC No. 40795) were prepared using a random primer kit (New England Biolabs, Inc.). The probes were used according to the method Benton & Davis (Maniatis, et al. *supra*) to screen the *Pyrococcus* genomic library using the hybridization conditions discussed above. About one per cent of the plaques were positive and ten positive plaques were picked and purified by reinfection and replating 3 times (until 90-100% of the plaques were positive for each isolate). Plate lysates (Maniatis, *supra*) of phage were prepared from each isolate and used to infect *E. coli* cultures. 0.1 ml of each plate lysate was mixed with 0.2 ml of cells (OD 600:2) The bacterial cells were harvested just before lysis and suspended in 0.05 M NaCl, 0.01M Tris (pH8.0), 0.1 mM EDTA, 0.1% Triton X-100 and 200 µg/ml lysozyme (3 volumes per volume of cells) and heated to 37°C for about 1 minute or until cell lysis occurred. The lysed extracts were immediately heated at 75°C for 30 minutes, centrifuged and the supernatant solution assayed for heat stable DNA polymerase activity, according to the method described above. Three of the ten isolates showed significant polymerase activity and the clone (B9) showing the most activity was investigated further.

[0067] The phage DNA was isolated from B9 and the insert DNA was examined by restriction enzyme digestion. Digestion with Sal I gave the expected two arms of λDASH plus a 15 Kb insert. Digestion with BamH I gave the two arms of λDASH plus three insert fragments of 7, 4.8 and 3 Kb. Each of these fragments were purified by agarose gel electrophoresis, eluted and ligated into the BamH I site of pUC19. The ligation mixtures were used to transform *E. coli* ER2207 which gives white colonies when plasmids contain an insert and blue colonies with no inserts on indicator agar media (X-gal plus IPTG). No white colony transformants were obtained with the 7 kb fragment. Three white colonies and twenty-seven blue colonies were obtained with the 4.8 Kb fragment and twenty white and twenty-one blue colony transformants were obtained with the 3 Kb fragment. All three 4.8 Kb white colony transformants expressed heat stable DNA polymerase activity. None of the transformants with the 3 Kb fragment expressed heat stable polymerase activity. The three clones carrying the 4.8 Kb *Pyrococcus* DNA fragment all had about the same specific activity for heat stable DNA polymerase and one was picked for further study (NEB#720). This clone designated NEB#720 was deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland, under the terms of the Budapest Treaty, on October 1, 1991 and bears ATCC No. 68723. NEB#720 yielded 1700 units of DNA polymer-

ase activity per gram of cells and was used for the large scale preparation of this enzyme.

[0068] A restriction endonuclease map of the 4.8 Kb BamH I fragment containing the *Pyrococcus* sp. DNA polymerase gene is shown in Figure 2. A partial DNA sequence of this gene is set forth in the Sequence Listing as SEQ ID NO:1. By comparison of the *Pyrococcus species* polymerase DNA and protein sequences of the present Invention with *Thermococcus litoralis* DNA polymerase DNA and protein it was discovered that at least one intervening sequence is present within the *Pyrococcus* DNA polymerase gene. The *Pyrococcus* intervening sequence is within the conserved pol α motif region III, and begins at nucleotide 1839 of SEQ ID NO:1.

EXAMPLE III

PURIFICATION OF RECOMBINANT PYROCOCCUS SPECIES DNA POLYMERASE

[0069] *E. coli* NEB#720 (ATCC No. 68723) was grown in a 25 liter fermentor in media containing 10 g/liter tryptone, 5 g/liter yeast extract, 5 g/liter NaCl and 100 mg/liter ampicillin at 37°C and induced with 0.3 mM IPTG at mid exponential growth phase and incubated an additional 4 hours. The cells are harvested by centrifugation and stored at -70°C.

[0070] 92 grams of cells were thawed and suspended in Buffer A (100 mM NaCl, 10 mM KPO₄ at pH 7.4, 0.1 mM EDTA, 0.1% Triton X-100 200 μ g/ml lysozyme) to a total volume of 350 ml. The cells were lysed by incubation at 37°C until the mixture became extremely viscous (about 5 min.). The crude extract was immediately heated to 75°C for 30 minutes. The denatured *E. coli* proteins were removed by centrifugation and the supernatant solution was used for the isolation of the *Pyrococcus* sp. heat stable DNA polymerase.

[0071] The supernatant solution was brought to 0.3M NaCl and passed through a DEAE-cellulose column 2x30cm prepared in a buchner funnel to remove nucleic acids and the flow through passed through an Affi-Gel Blue chromatography column 4x10cm (125 ml) and washed with 300 ml 0.3M NaCl, 0.01M KPO₄ (pH 7.4) 0.1mM EDTA and then eluted with a 1500 ml linear gradient of NaCl from 0.3M to 2.0M in the same buffer.

[0072] The column fractions were assayed for DNA polymerase activity. Briefly, 1-4 μ l of fractions were incubated for 5-10 minutes at 75°C in 50 μ l of DNA polymerase buffer (10 mM KCl, 20 mM Tris-HCl (pH 8.8 at 24°C), 10 mM (NH₄)₂SO₄, 2 mM MgSO₄ and 0.1% Triton X-100) containing 30 μ M each dNTP and ³H-labelled TTP, 0.5 mg/ml activated calf thymus DNA and 100 μ g/ml acetylated BSA. The assay mixtures were applied to Whatman 3 mm filters and the filters were subjected to three washes of 10% TCA followed by two washes of cold ethanol. After drying of the filters, bound radioactivity representing incorporation of ³H-TTP into the DNA was measured. The active fractions were pooled dialyzed against 0.1M NaCl, 0.01M KPO₄ (pH7.4), 0.1mM EDTA and then passed through a phosphocellulose column 4x12cm (150 ml) and the column washed with 300 ml of the same buffer then eluted with a linear gradient of NaCl from 0.1M to 1.5M. Active fractions were pooled and diluted with an equal volume of H₂O and passed through a 1.0 ml Pharmacia HPLC Mono Q column and eluted with a 60 ml linear gradient of NaCl from 0.05M to 1.0M. Active fractions were pooled and stored at -20°C. The *Pyrococcus* sp. DNA polymerase was approximately 50% pure at this stage as determined by visual assessment of a Coomassie Blue stained gel and had a specific activity for DNA synthesis of 70,000-100,000 units per mg, and represented 8% of enzyme activity present in the crude extracts.

The purified polymerase was substantially free of contaminating DNA and contaminating nucleases.

[0073] The DNA polymerase activity corresponded to major 92-97,000 Kd Coomassie Blue staining band in Figure 3. This was determined in the following manner. After electrophoresis in SDS 10-20% polyacrylamide gel and before staining with Coomassie Blue, the gel was soaked in 1.0% Triton X-100, 0.01M Tris HCl (pH7.4) overnight at 4°C with 3 changes of 200 ml each to remove sodium dodecylsulfate (SDS) from the gel and replace it with the non-ionic detergent Triton X-100. After the last wash, the gel was soaked in 0.1% Triton X-100, 0.01M Tris HCl (pH7.4) for 2 hours and then dried briefly with paper towel and placed on a glass plate. The gel was covered with a piece of Whatman No. 54 filter paper and 0.3 ml of assay buffer containing ³²P-d CTP (10-20x10⁶ cpm per μ l) added to the center of the filter paper. After the liquid had diffused towards the sides of the paper, a second glass plate was added on top of the filter paper and the glass-gel-paper-glass sandwich tightly taped together and incubated at 75°C for 60 minutes. The paper was removed from the gel, washed three times (30 minutes each wash) with 10% TCA containing 1mM dCTP and twice with isopropanol. The paper was air dried and exposed to an X-ray film overnight. When developed, a black spot on the film could be superimposed on the 92-97,000 Kd Coomassie Blue band on the stained gel indicating that the major band on the gel in Figure 3 is the DNA polymerase.

[0074] Western blot analysis (Towbin, et al, *PNAS* (1979) 76:4350-4354) of the purified recombinant protein using antibody prepared against the closely related DNA polymerase from *Thermococcus litoralis*, indicated a major band at 92,000-97,000 Kd identical to the Coomassie Blue stained band and at approximately the same location on the gel (Figure 4).

EXAMPLE IV

DETERMINATION OF 3'-5' PROOFREADING ACTIVITY

5 1. Response of *T. litoralis* DNA Polymerase to the Absence of Presence of Deoxynucleotides

[0075] The levels of exonuclease activities associated with polymerases show very different responses to deoxynucleotides. Nonproofreading 5'-3' exonucleases are stimulated tenfold or greater by concomitant polymerization afforded by the presence of deoxynucleotides, while proofreading 3'-5' exonucleases are inhibited completely by concomitant polymerization. Lehman, I.R. *ARB* (1967) 36:645.

[0076] The *Pyrococcus* sp. DNA polymerase or polymerases with well-characterized exonuclease functions (T4 Polymerase, Klenow fragment) were incubated with 1 µg 3 H-thymidine-labelled double-stranded DNA (10^5 CPM/µg) in polymerization buffer (70 mM tris pH 8.8 at 24°C), 2 mM MgCl₂, 0.1% Triton and 100 µg/ml bovine serum albumin. After an incubation period of three hours (experiment 1) or four hours (experiment 2) at either 70°C (thermophilic polymerases) or 37°C (mesophilic polymerases), the exonuclease-hydrolyzed bases were quantified by measuring the acid-soluble radioactively-labelled bases.

[0077] As shown in Table 1, the Taq DNA polymerase, with its 5'-3' exonuclease activity, shows stimulation of exonuclease activity when deoxynucleotides were present at 30 µM. However, polymerases with 3'-5' proofreading exonuclease activities, such as the T4 polymerase, Klenow fragment of *E. coli* polymerase I, *T. litoralis* DNA polymerase or the *Pyrococcus* DNA polymerase showed the reverse, an inhibitory response to the presence of deoxynucleotides.

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TABLE 1

Acid Soluble CPM (Exonuclease Activities)*

Amount	Type of DNA Polymerase	no dNTPS	30uM dNTPS	Effect Upon Adding NTPS
5 units	Taq Polymerase	338	2539	8X increase
5 units	T4 Polymerase	*46001	10418	>4X decrease
5 units	Klenow Fragment of <i>E. coli</i> Pol. 1	8757	408	22X decrease
5 units	<i>T. litoralis</i> Polymerase	8573	795	11X decrease
5 units	<i>Pyrococcus</i> sp. Polymerase	9798	1411	7X decrease

* Nonlinear Range of Assay

2. Comparison of 3'-5' Proofreading Activity of *Pyrococcus* sp. DNA Polymerase to *Thermococcus litoralis* DNA Polymerase

5 [0078] To further characterize the 3'-5' exonuclease activity of *Pyrococcus* sp. DNA polymerase, both enzymes were compared at the same DNA polymerization units for 3'-5' proofreading exonuclease activity (Figure 5). The results indicate that at equal DNA polymerization units, *Pyrococcus* sp. DNA polymerase has 2.5 times the exonuclease activity as *Thermococcus litoralis* DNA polymerase. This would suggest greater fidelity during DNA synthesis.

10 EXAMPLE V

PYROCOCCUS SP. DNA POLYMERASE HALF LIFE DETERMINATION

15 [0079] The thermostability or half-life of the *Pyrococcus* sp. DNA polymerase purified as described above in Example III was determined by the following method. Purified recombinant *Pyrococcus* sp. DNA polymerase (40 units/ml) was preincubated at 95°C or 100°C in reaction buffer lacking dNTPs and DNA (reaction buffer 10 mM KCl, 10 mM (NH₄)₂ SO₄, 20 mM Tris-HCl (pH 8.8 at 25°C), 2 mM MgSO₄, 0.1% TRITON X-100, supplemented with 0.1 mg/ml BSA). At times indicated in Figure 6, an aliquot of the enzyme mixture was diluted four-fold into reaction buffer at 70°C which contained [³H] labelled dNTPs and primed M13 DNA substrate (final concentration 0.2 mM and 20nM, respectively) and the initial rate of [³H] incorporation into acid insoluble material was monitored. Activity is expressed relative to activity present prior to treatment at 95°C or 100°C.

20 [0080] As shown in Figure 6, the half life of the *Pyrococcus* sp. DNA polymerase at 95°C was 23 hours, and 8 hours at 100°C.

25 SEQUENCE LISTING

[0081]

(1) GENERAL INFORMATION:

30 (i) APPLICANT:

35 COMB, DONALD G.
PERLER, FRANCINE
KUCERA, REBECCA
KONG, HUIMIN
JACK, WILLIAM F.

40 (ii) TITLE OF INVENTION: PURIFIED THERMOSTABLE DNA POLYMERASE OBTAINABLE FROM PYROCOCCUS SPECIES

(iii) NUMBER OF SEQUENCES: 2

(v) COMPUTER READABLE FORM:

45 (A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

50 (A) APPLICATION NUMBER:

(B) FILING DATE:

(C) CLASSIFICATION:

55 (2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3420 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: not relevant

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

10 GGATCCCTCT CTTTTGGTA ACCCCATAAG TCATTCCCTC AACCAAAACT TCAGCATCGT 60
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 20 TGTATCTCCA CATTCTAACT AACGCTCCAG GCCCAGGATC AACGTAGATG TTTTGCTCG 180
 25 CCTTAATGAA GAAGCCACCA GTGGCTCTTG CCTGCCTTAT CGTGACGAAC CTTCCACCAC 240
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 65 TAAGAGAGCA TTCCGCAGTT ATTGACATCT TTGAGTACGA CATTCCGTTC GCGAAGAGGT 720
 70 ACCTAATAGA CAAAGGCCTA ATTCCAATGG AAGGCGATGA AGAGCTCAAG TTGCTCGCAT 780
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10 (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 5837 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS:
(D) TOPOLOGY: not relevant

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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5 GAATTCGCGA TAAAATCTAT TTTCTTCCTC CATTTCCTAA TTTCAAAAAC GTAAGCATGA 60
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 40 GGGAGTTCTT GGGATCC 5837

Claims

45 1. A purified thermostable enzyme obtainable from *Pyrococcus sp.* which catalyzes the polymerization of DNA, wherein said enzyme is obtainable from *E.coli* NEB#720 (ATCC # 68723).

50 2. The thermostable enzyme of claim 1, having a molecular weight of about 92,000 to 97,000 daltons.

3. The thermostable enzyme of claim 1, having a 3'-5' exonuclease activity.

4. The thermostable enzyme of claim 3, wherein the 3'-5' exonuclease activity is inactivated.

55 5. The thermostable enzyme of claim 1, having a half life of about 8 hours at 100°C.

6. The thermostable enzyme of claim 1, having a half life of about 23 hours at 95°C.

7. An isolated DNA sequence coding for the thermostable enzyme of claim 1.
8. A vector containing the DNA sequence of claim 7.
- 5 9. A microbial host transformed with the vector of claim 8.
10. The transformant of claim 9, wherein said transformant is *E. coli* NEB#720 (ATCC No. 68723).
11. A process for the preparation of *Pyrococcus sp.* DNA polymerase comprising culturing the transformed microbial host of any of the claims 9 or 10 under conditions suitable for the expression of *Pyrococcus sp.* DNA polymerase and recovering *Pyrococcus sp.* DNA polymerase.
12. The DNA of claim 7, wherein the DNA contains an intervening DNA sequence.
- 15 13. A process according to claim 11 for increasing expression of *Pyrococcus sp.* DNA polymerase comprising:
 - (a) deleting an intervening DNA sequence from DNA according to claim 7;
 - (b) introducing the DNA of step (a) into an expression vector;
 - (c) transforming a microbial host with the expression vector of step (b);
 - 20 (d) culturing the transformed host of step (c), under conditions suitable for the expression of *Pyrococcus sp.* DNA polymerase; and
 - (e) recovering the polymerase.

25 **Patentansprüche**

1. Gereinigtes thermostabiles Enzym, erhältlich von *Pyrococcus sp.*, das die Polymerisation von DNA katalysiert, wobei das genannte Enzym von *E. coli* NEB Nr. 720 (ATCC Nr. 68723) erhältlich ist.
- 30 2. Thermostabiles Enzym nach Anspruch 1, mit einer relativen Molekülmasse von etwa 92.000 bis 97.000 Dalton.
3. Thermostabiles Enzym nach Anspruch 1, mit einer 3'-5'-Exonucleaseaktivität.
4. Thermostabiles Enzym nach Anspruch 3, wobei die 3'-5'-Exonucleaseaktivität inaktiviert ist.
- 35 5. Thermostabiles Enzym nach Anspruch 1, mit einer Halbwertszeit von etwa 8 Stunden bei 100°C.
6. Thermostabiles Enzym nach Anspruch 1, mit einer Halbwertszeit von etwa 23 Stunden bei 95°C.
- 40 7. Isolierte DNA-Sequenz, die das thermostabile Enzym aus Anspruch 1 kodiert.
8. Vektor, der die DNA-Sequenz von Anspruch 7 enthält.
9. Mikrobieller Wirt, transformiert mit dem Vektor aus Anspruch 8.
- 45 10. Transformante nach Anspruch 9, wobei die genannte Transformante *E. coli* NEB Nr. 720 (ATCC Nr. 68723) ist.
11. Verfahren zur Herstellung von *Pyrococcus sp.* DNA-Polymerase, umfassend das Kultivieren des transformierten mikrobiellen Wirts nach einem der Ansprüche 9 oder 10 unter Bedingungen, die für die Expression von *Pyrococcus sp.* DNA-Polymerase und Wiedergewinnung von *Pyrococcus sp.* DNA-Polymerase geeignet sind.
- 50 12. DNA nach Anspruch 7, wobei die DNA eine Intervening-DNA-Sequenz enthält.
13. Verfahren nach Anspruch 11 für die Steigerung der Expression von *Pyrococcus sp.* DNA-Polymerase, umfassend die folgenden Schritte:
 - (a) Deleteieren einer Intervening-DNA-Sequenz von DNA nach Anspruch 7;
 - (b) Einleiten der DNA aus Schritt (a) in einen Expressionsvektor;

(c) Transformieren eines mikrobiellen Wirts mit dem Expressionsvektor von Schritt (b);
(d) Kultivieren des transformierten Wirts aus Schritt (c) unter Bedingungen, die für die Expression von *Pyrococcus sp.* DNA-Polymerase geeignet sind; und
(e) Wiedergewinnen der Polymerase.

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Revendications

1. Enzyme thermostable purifiée pouvant être obtenue de *Pyrococcus sp.* qui catalyse la polymérisation de l'ADN, où ladite enzyme peut être obtenue d'*E. coli* NEB#720 (ATCC # 68723).
- 10 2. Enzyme thermostable de la revendication 1, ayant un poids moléculaire d'environ 92.000 à 97.000 daltons.
3. Enzyme thermostable de la revendication 1, ayant une activité exonucléase 3'-5'.
- 15 4. Enzyme thermostable de la revendication 3, dans laquelle l'activité exonucléase 3'-5' est inactivée.
5. Enzyme thermostable de la revendication 1, ayant une demi-vie d'environ 8 heures à 100°C.
- 20 6. Enzyme thermostable de la revendication 1, ayant une demi-vie d'environ 23 heures à 95°C.
7. Séquence d'ADN isolée codant l'enzyme thermostable de la revendication 1.
- 25 8. Vecteur contenant la séquence d'ADN de la revendication 7.
9. Hôte microbien transformé avec le vecteur de la revendication 8.
10. Le transformant de la revendication 9, où ledit transformant est *E. coli* NEB#720 (ATCC n° 68723).
- 30 11. Procédé pour la préparation d'ADN polymérase *Pyrococcus sp.* comprenant cultiver l'hôte microbien transformé de l'une quelconque des revendications 9 ou 10 dans des conditions qui conviennent à l'expression de l'ADN polymérase *Pyrococcus sp.* et récupérer l'ADN polymérase *Pyrococcus sp.*
12. L'ADN de la revendication 7, où l'ADN contient une séquence d'ADN interposée.
- 35 13. Procédé selon la revendication 11 pour augmenter l'expression de l'ADN polymérase *Pyrococcus sp.* comprenant :
 - (a) éliminer une séquence d'ADN interposée de l'ADN selon la revendication 7;
 - (b) introduire l'ADN de l'étape (a) dans un vecteur d'expression;
 - 40 (c) transformer un hôte microbien avec le vecteur d'expression de l'étape (b);
 - (d) cultiver l'hôte transformé de l'étape (c), dans des conditions qui conviennent à l'expression de l'ADN polymérase *Pyrococcus sp.*; et
 - (e) récupérer la polymérase.

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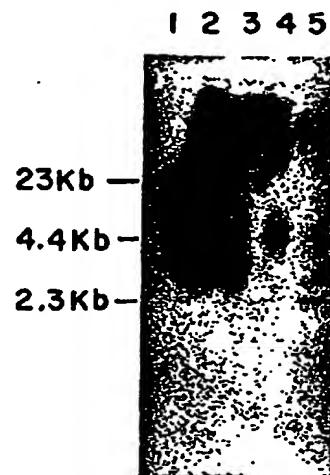
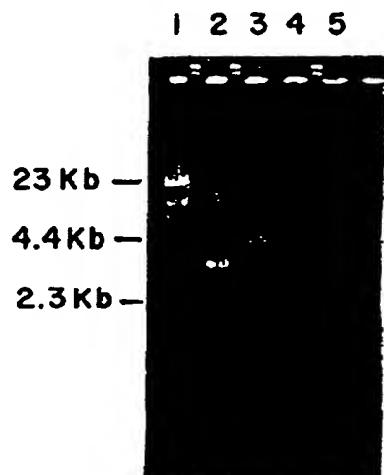


FIG. 1A

FIG. 1B

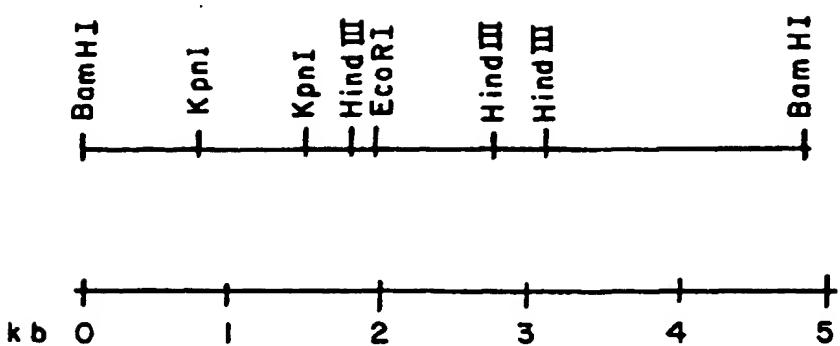
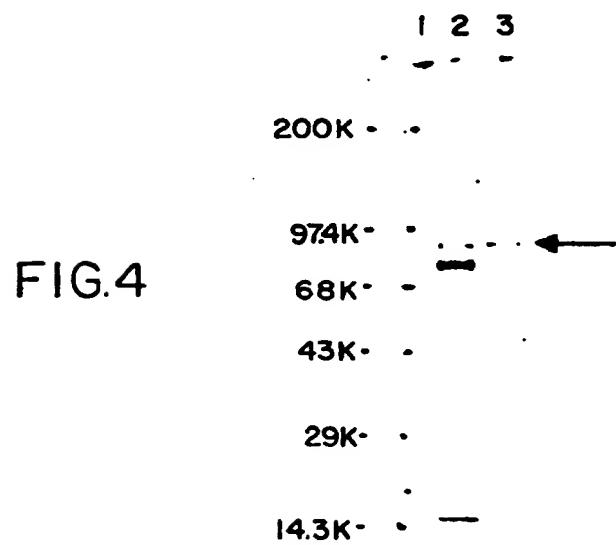


FIG. 2



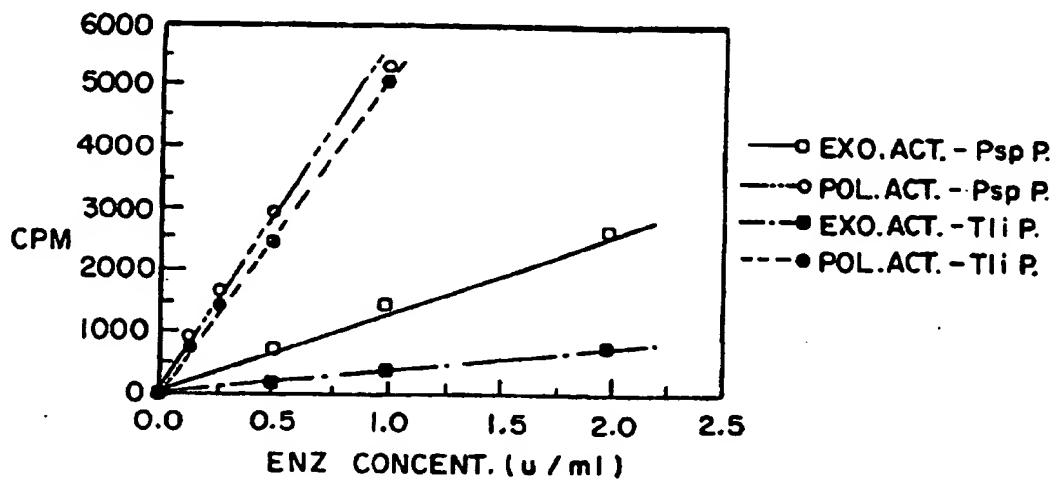


FIG. 5

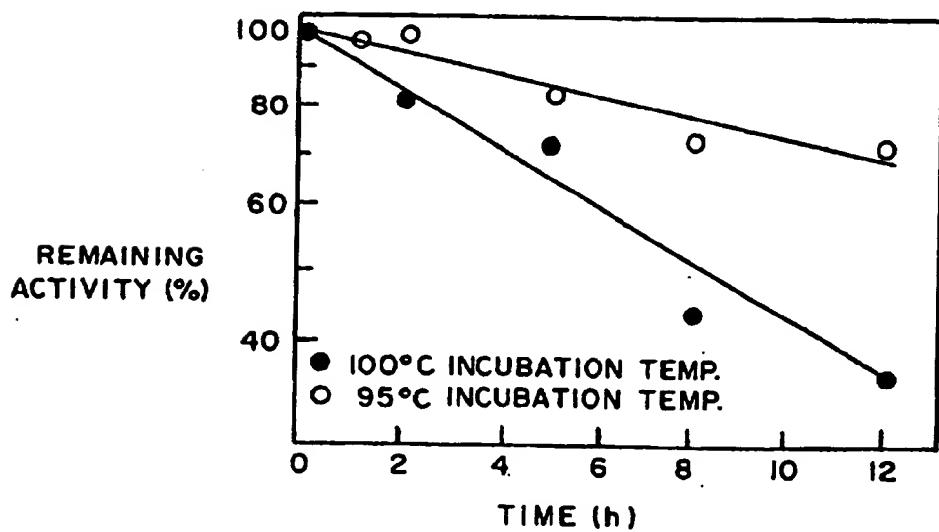


FIG. 6